



# DTI JadeAmp FabTaq Premix

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Instruction Manual for use



User Manual for DTI JadeAmp FabTaq premix

Catalogue number: DT0201.80, DT0201.320

*Note: Applicable to all pack sizes*

Manufactured by  
DSS Takara Bio India Pvt Ltd  
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**1) Product Description:**

DTI JadeAmp FabTaq premix is a 2X premix composed of a DNA polymerase, optimized reaction buffer, dNTPs, and a density reagent. The premix also contains a vivid green dye that will separate into blue and yellow dye fronts when run on an agarose gel. The premix simplifies PCR assembly; simply add primers, template, and water and start the reaction. After PCR, the reaction mixture can be applied directly to a gel for analysis.

**2) Storage:** -20 °C (or 4 °C for 3 months)

If the premix will be used frequently, store at 4°C; repeated freezing and thawing will decrease activity. Mix the premix gently and briefly centrifuge before use.

**3) Applications:**

DNA amplification by PCR  
Colony PCR  
PCR screening

**4) Quality Control Data**

Please see the Certificate of Analysis (CoA) for each lot.

**5) Components (25 µl per reaction)**

Components	Cat# DT0201.80 (80 reactions)	Cat# DT0201.320 (320 reactions)
DTI JadeAmp FabTaq premix	1 ml	1 ml x 4
dH2O	1 ml	1 ml x 4

**6) Materials required but not provided**

**Reagents:** dNTP, PCR primers, sterile purified water, template

**Equipment:** Thermal cycler (DTI FabSpeed thermal cycler, model# TCST-9622)

**Consumables:** PCR tubes, Micropipettes and tips

**7) Protocol****General Reaction Composition for PCR:**

Reagent	Volume	Volume
DTI JadeAmp FabTaq premix (2X Premix)	12.5 µl	25 µl
Template	< 500 ng	< 500 ng
Forward Primer	0.1 µM (final conc.)	0.2 µM (final conc.)
Reverse Primer	0.1 µM (final conc.)	0.2 µM (final conc.)
dH2O	up to 25 µl	up to 50 µl
<b>Total</b>	<b>25 µl</b>	<b>50 µl</b>

**8) Suggested PCR Conditions:****3 Step (up to 6 kb)**

98°C	10 sec	} 30 cycles
60°C*	30 sec	
72°C	1 min/kb	

**2 Step (over 6 kb)**

98°C	10 sec	} 30 cycles
68°C	1 min/kb	

\* For optimal results, primers should have a  $T_m > 60^\circ\text{C}$ . The following formula is commonly used for estimating the  $T_m$  of the primers.

$$T_m (\text{°C}) = [(n_A + n_T) \times 2] + [(n_G + n_C) \times 4] - 5$$

n : the number of adenine (A), thymidine (T), guanidine (G), or cytosine (C) bases in primer

NOTE: Denaturation conditions vary depending on the thermal cycler and tubes used for PCR. Denaturation for 5 - 10 sec at 98°C or 20 - 30 sec at 94°C is recommended.

**9) PCR product:**

PCR products generated with DTI JadeAmp FabTaq premix have a single A at the 3'-termini, and PCR products can be directly cloned into a T-vector. It is also possible to clone products into blunt-end vectors after blunting and phosphorylation of the ends.

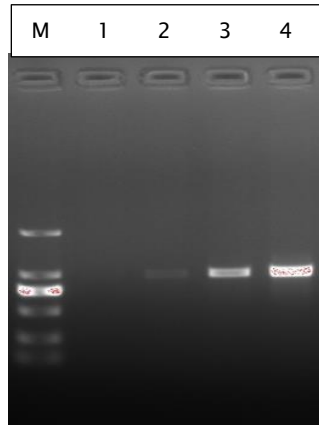
**10) Dye marker migration:**

When 5  $\mu\text{l}$  of the reaction mixture is used for electrophoresis on a 1% DTI agarose routine (cat# DT1701.500) gel, the blue dye front migrates near 3 - 5 kb and the yellow dye front is below 50 bp.

**11) Experimental sample:**

In the below study, the DTI JadeAmp FabTaq premix is capable of amplifying 1 kb fragment from 1ng, 10 ng and 100 ng of human genomic DNA as template. The protocol used for the assay and the results are as follows:

98 C	10 sec	} 30 Cycles
60 C	30 sec	
72 C	1 min	



Lane M: 1 kb DNA marker  
Lane 1: Test – 100 pg  
Lane 2: Test– 1 ng  
Lane 3: Control - 10 ng  
Lane 4: Control - 100 ng

Figure 1: Gel electrophoresis image of PCR

Visit <https://store.dsstakarabio.com/pages/dti-jadeamp-fabtaq-premix> for more detailed product information






For more information contact directly below;

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#### Description of Symbol Used:

-  Catalogue number
-  Batch Code
-  Date of Manufacturing
-  Use-by-date
-  Contains sufficient for <n> tests